

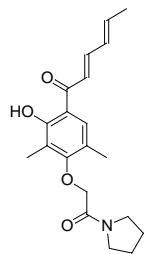
Supporting Information: Pettus, T. R. R. *et al*

Selected physical properties and spectra of compounds **9**, **12**, **13**, **14**, **16** and **17** and synthetic **1** and **2** are provided.

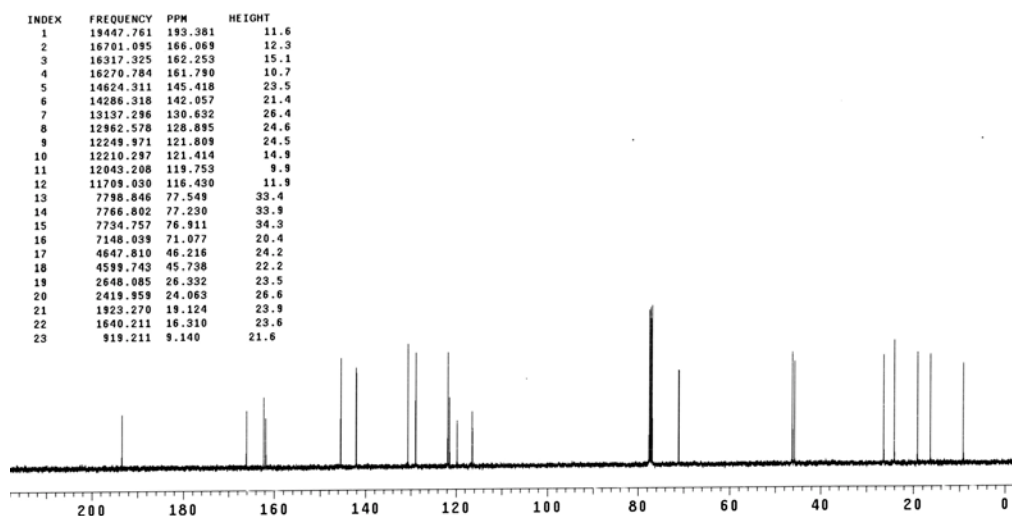
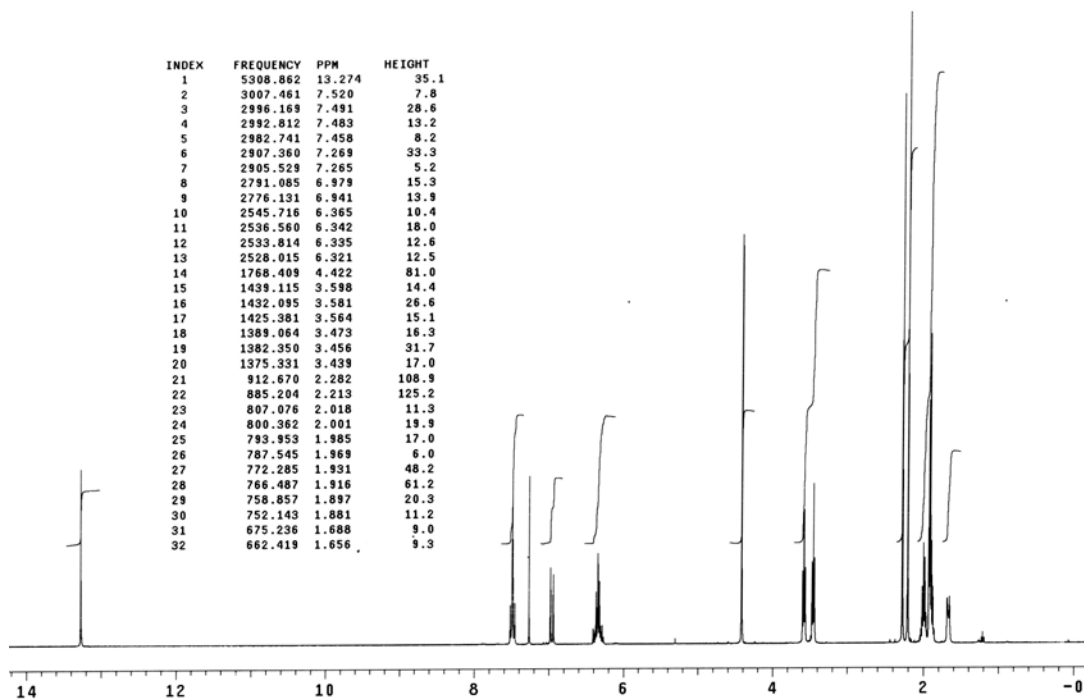
General information: The following section provides general information. Immediately following this information are general and/or specific reaction conditions and spectral data for compounds along with a representative ^1H NMR spectrum.

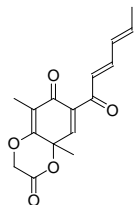
Reactions were monitored by analytical thin-layer chromatography on EM-Science hard layer silica gel-60^{F-250} plates cut into 1x2.5^{cm} sections. Visualization was effected by ultraviolet light (254 nm), followed by dipping the plate into the appropriate stain and charring on a *Fisher*[®] micro-hot plate. The primary stain was made with [25 g of phosphoromolybdic acid and 10 g of cerium sulfate and 60 mL H_2SO_4 and 940 mL of H_2O]. Distillations were accomplished with a short-path condenser configured with multiple receiving flasks. A specific low pressure (760-1 mm Hg) was obtained and monitored with a *Buchi*[®]-vacuum controller model number B-721 in combination with a *Welch*[®] direct drive pump model number 8915A. Lower pressures were achieved using a vacuum manifold connected to an oil-diffusion pump and backed by a *Welch*[®] direct drive vacuum pump model number 8910A. Chromatography was performed following the method prescribed by W. C. Still.¹ In reactions, where water was *not* present as a solvent, reagent or by-product, vessels were flame-dried under a slow nitrogen flow. A slight positive pressure of dry nitrogen was maintained via rubber septa seal during the course of the reaction. The nitrogen stream originated from a high pressure 55 L (l) N_2 tank and was further dried by passing through a Drierite[®] tube. Reagents were purified according to the procedures describe in the *Perrin & Perrin* laboratory manual. All reactions were stirred with Teflon-coated magnetic stir bars via *Thomas*[®] magnetic stirrers. Removal of solvents was normally accomplished using a *Buchi*[®] rotary evaporator model number R-114 connected to a KNF[®]-vacuum pump model number UN820-3. The condenser was cooled to 0 °C by a *Fisher*[®] circulator bath model number 1013S. Trace solvents were removed by a *Labconco*[®] freeze dryer system at a pressure of approximately 0.01 mm Hg. The following solvents were distilled before use, under a slight positive pressure of nitrogen. Diethyl ether was distilled from sodium benzophenone ketyl. Methylene chloride, di-*n*-butyl ether and acetone were distilled from calcium hydride. The chloroform, isopropyl alcohol and hexanes used for infrared spectra and HPLC analyses. Acetonitrile was distilled over CaH_2 for circular dichroism spectra. Melting points were determined on a *Fisher-Johns*[®] melting point apparatus and are uncorrected. ^1H -NMR spectra were recorded at 400 MHz or 500 MHz on a *Varian*[®] spectrometer. Chemical shifts are reported in ppm from tetramethylsilane with the solvent resonance of CDCl_3 (7.27 ppm) as the internal standard. Chemical ^{13}C NMR spectra were recorded at (100 MHz) with a solvent resonance of CDCl_3 (77.23 ppm) as the internal standard. Infrared spectra were recorded on a *Shimadzu*[®] FT infrared spectrometer with 2 cm resolution in a solution cell (CH_2Cl_2). Infrared frequencies are reported in reciprocal centimeters (cm^{-1}). HPLC analyses for *enantiomeric excess* were performed isochratically using a *Shimadzu*[®] system (a 10 mL/min pump and dual channel UV detector) with a Chiracel columns. Non-enantiomeric separations of materials by HPLC used 5 μ 25 cm long spherisorb columns supplied by *Jones*[®] Chromatography. Circular dichroism spectra were recorded on an Aviv CD spectrophotometer model 202 at 25 °C. Optical rotations were recorded on a Perkin Elmer Polarimeter model 341 at 25 °C. Mass spectra were recorded on a VG-7035 mass spectrometer at an ionizing voltage of either 70 or 20 eV.

¹ Still, W. C.; Kahn, K.; Mitra, A. *J. Org. Chem.* **1978**, *43*, 2923-2924.

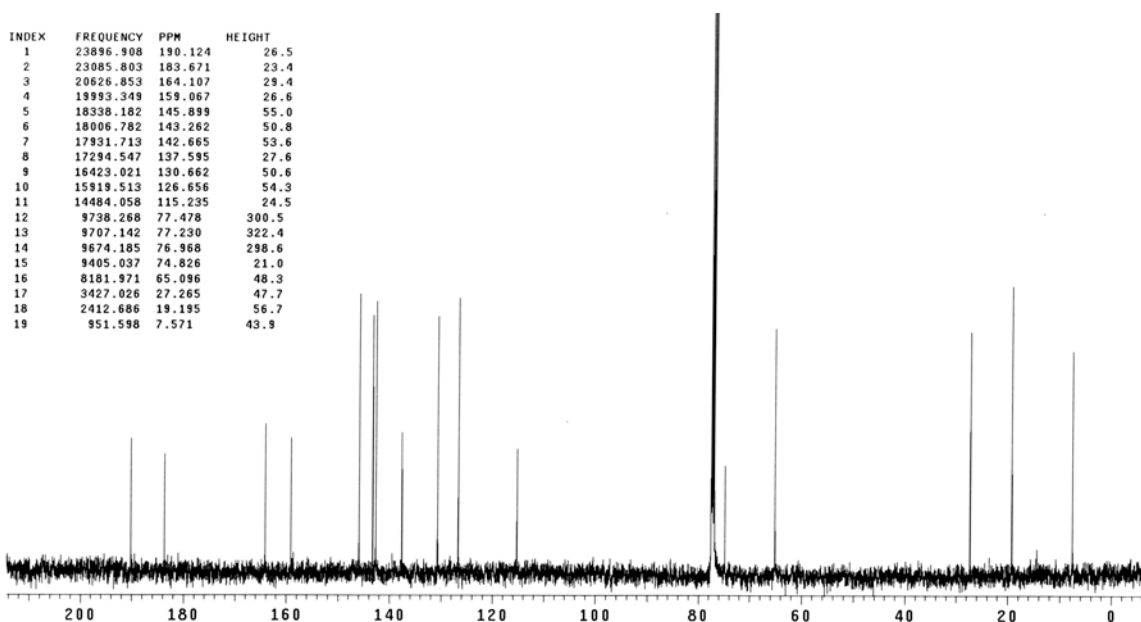
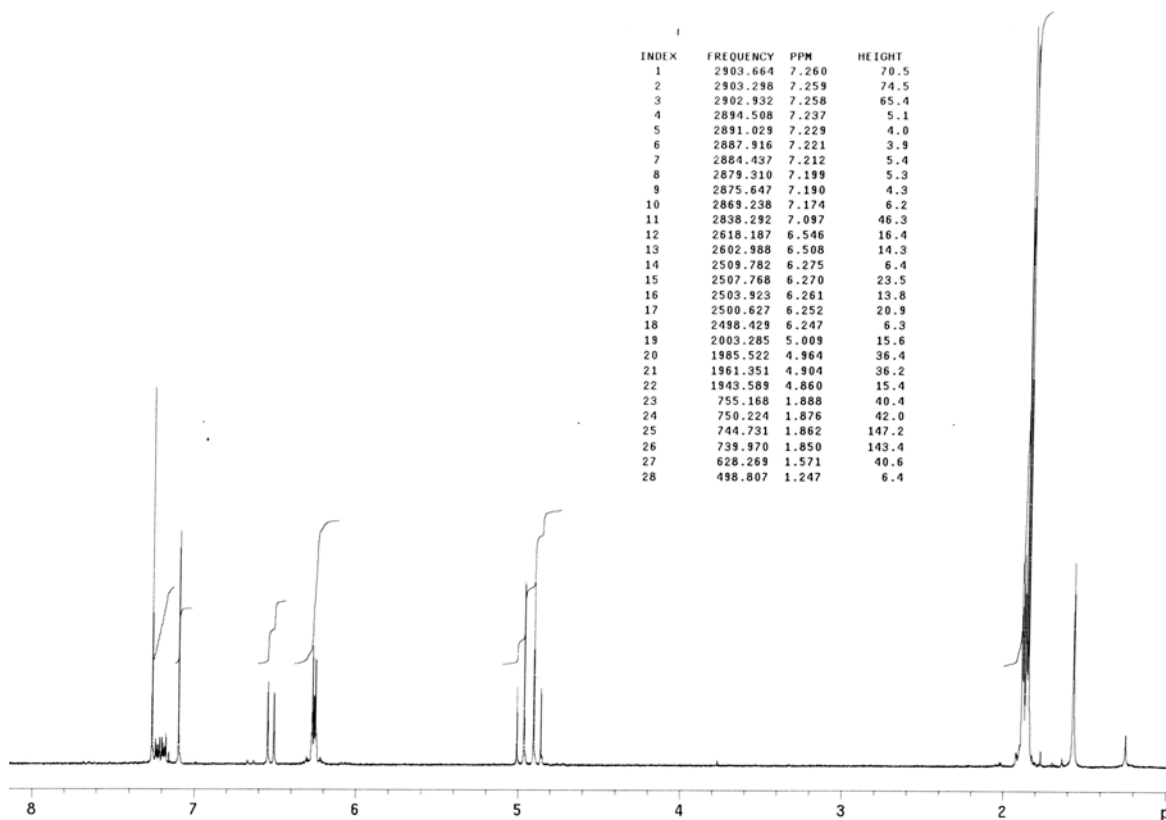


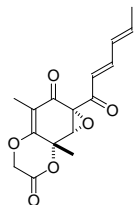
Amide 12: yellow oil $R_f=0.44$ $\text{CH}_2\text{Cl}_2/\text{acetone}$ (12/1); ^1H NMR (CDCl_3 , 400 MHz) δ 13.3 (s, OH), 7.50 (m, 2H), 6.96 (d, 1H, $J = 15.1$ Hz), 6.35 (m, 2H), 4.43 (s, 2H), 3.59 (t, 2H, $J = 7.1$ Hz), 3.46 (t, 2H, $J = 7.1$ Hz), 2.29 (s, 3H), 2.22 (s, 3H), 2.01 (m, 2H), 1.92 (d, 3H, $J = 6.2$ Hz), 1.90 (m, 2H); ^{13}C NMR (CDCl_3 , 100.6 MHz) δ 193.4, 166.1, 162.3, 161.8, 145.5, 142.1, 130.7, 128.9, 121.8, 121.5, 119.8, 116.5, 71.1, 46.3, 45.8, 26.4, 24.1, 19.2, 16.3, 9.2; IR (CHCl_3 , $\lambda\text{-max}$ cm^{-1}) 3688, 3009, 1644, 1500, 1361, 768; HRMS(EI) calcd for $\text{C}_{20}\text{H}_{25}\text{NO}_4$ 343.1784, found: 343.1783.



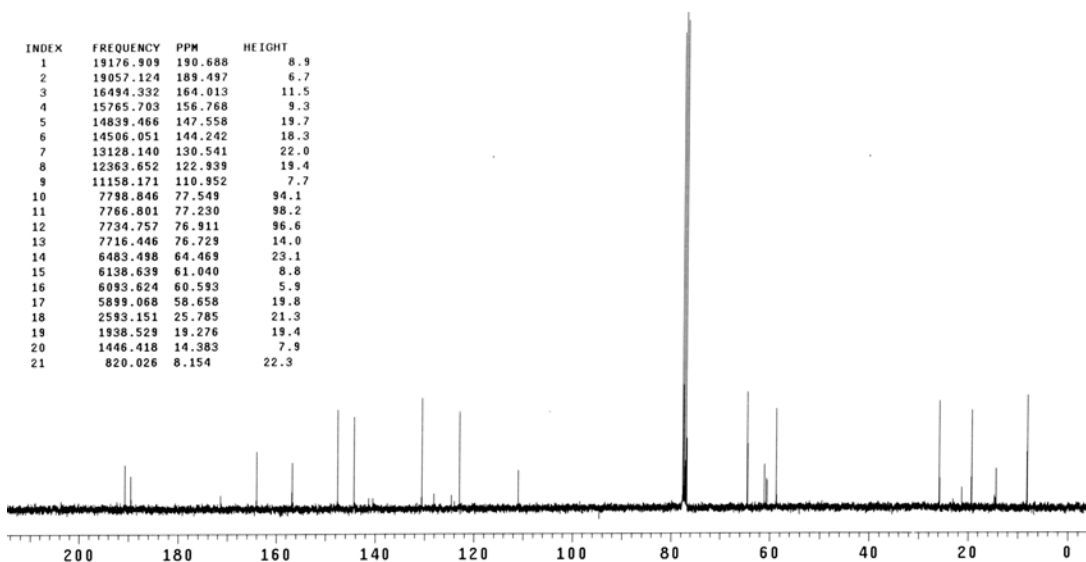
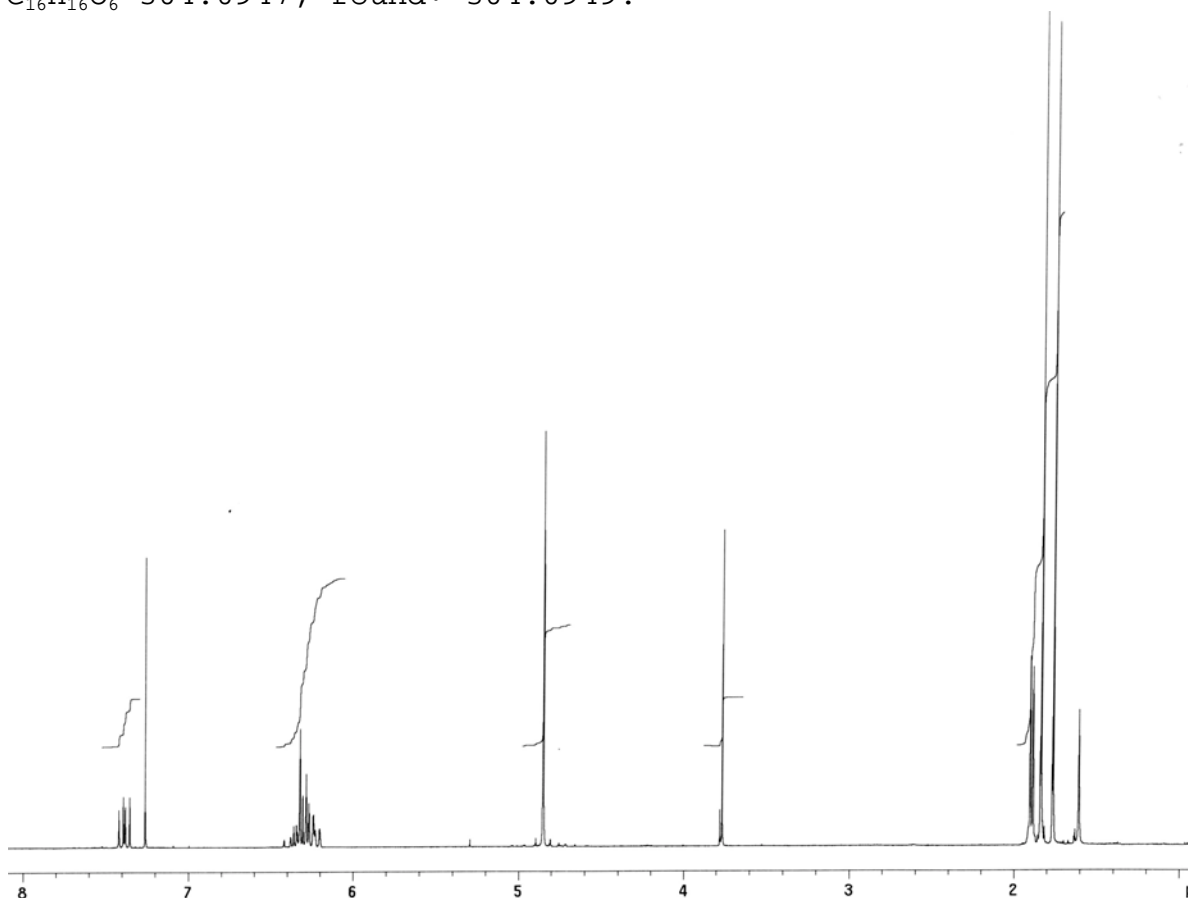


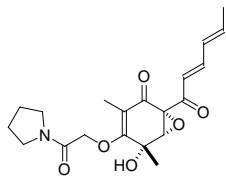
Dienone 9: pale yellow oil R_f=0.32 Hexanes/EtOAc (2/1); ¹H NMR (CDCl₃, 400 MHz) δ 7.20 (m, 1H), 7.09 (s, 1H), 6.52 (d, 1H, J = 15.2 Hz), 6.25 (m, 2H), 4.92 (AB, 2H, J = 17.8 Hz), 1.88 (d, 3H, J = 5.2 Hz), 1.86 (s, 3H), 1.84 (s, 3H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 190.1, 183.7, 164.1, 159.1, 145.9, 143.3, 142.7, 137.6, 130.7, 126.7, 115.2, 74.8, 65.1, 27.3, 19.2, 7.8; IR (CHCl₃, λ-max cm⁻¹) 3022, 1769, 1621, 1293, 1190; HRMS(CI) calcd for (M+H)⁺ C₁₆H₁₇O₅ 289.0998, found: 289.1070.



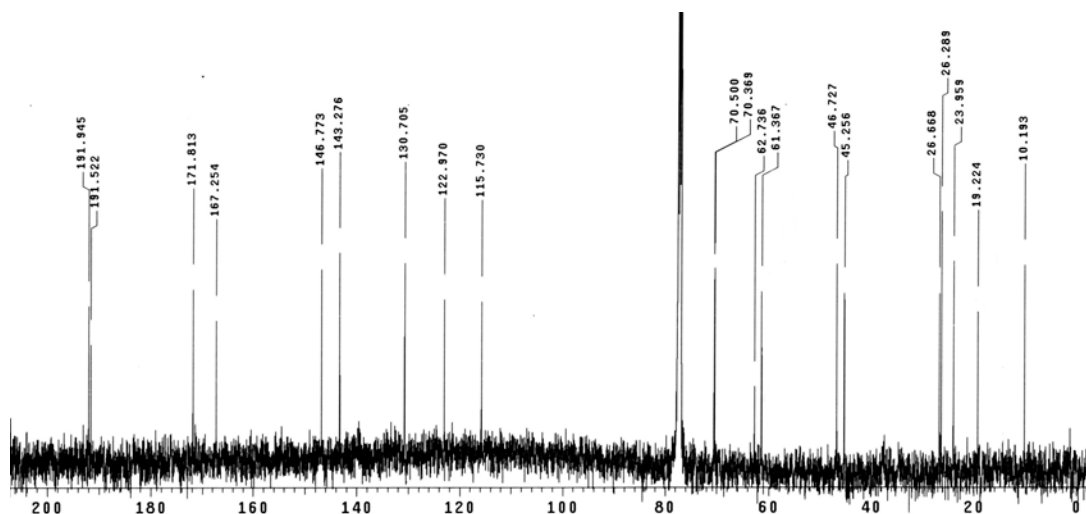
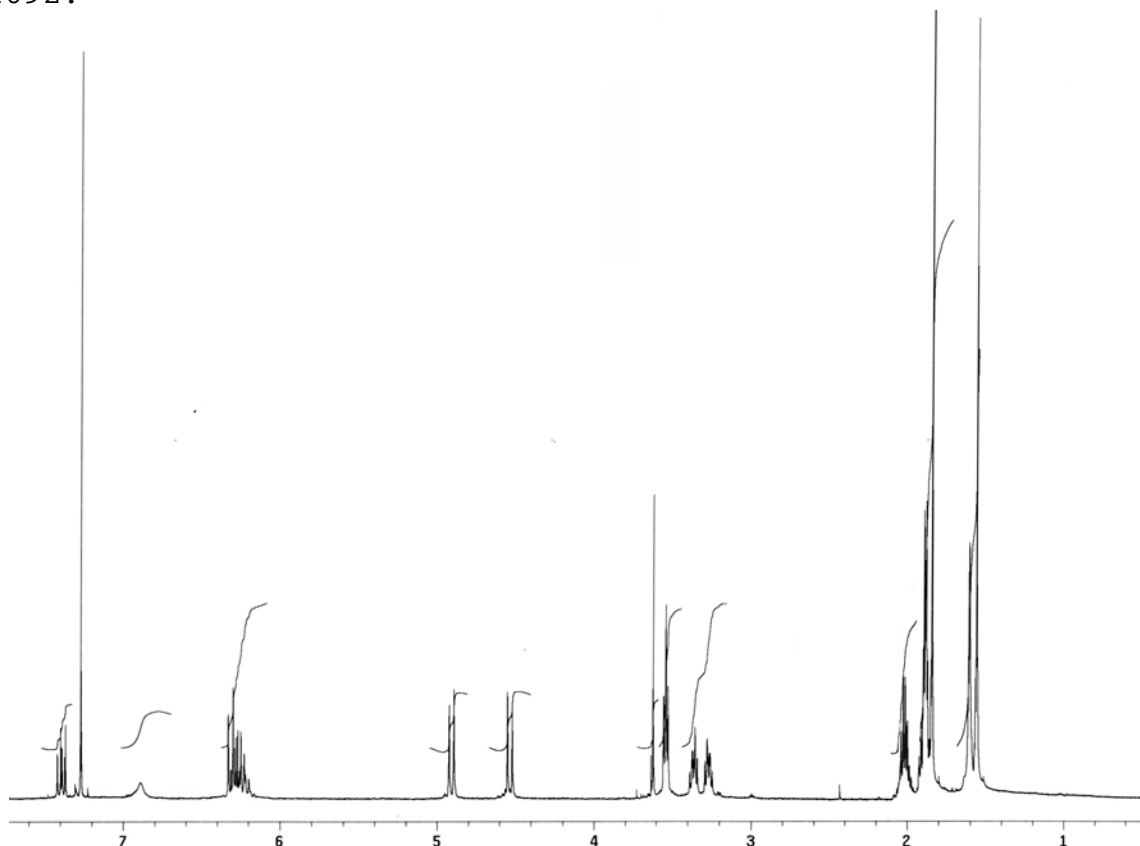


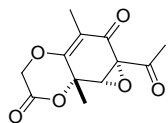
Epoxide 13: pale yellow oil $R_f=0.24$ Hexanes/EtOAc (2/1); ^1H NMR (CDCl_3 , 400 MHz) δ 7.38 (dd, 1H, $J_1 = 15.6$ Hz, $J_2 = 10.7$ Hz), 6.40–6.24 (m, 3H), 4.85 (s, 2H), 3.76 (s, 1H), 1.89 (d, 3H, $J = 6.77$ Hz), 1.83 (s, 3H), 1.76 (s, 3H); ^{13}C NMR (CDCl_3 , 100.6 MHz) δ 190.7, 189.5, 164.0, 156.8, 147.5, 144.2, 130.5, 122.9, 110.9, 70.7, 64.5, 61.0, 58.6, 25.8, 19.3, 8.2; IR (CHCl_3 , $\lambda\text{-max cm}^{-1}$) 3022, 1775, 1687, 1645, 1593, 1292; HRMS(EI) calcd for $\text{C}_{16}\text{H}_{16}\text{O}_6$ 304.0947, found: 304.0949.



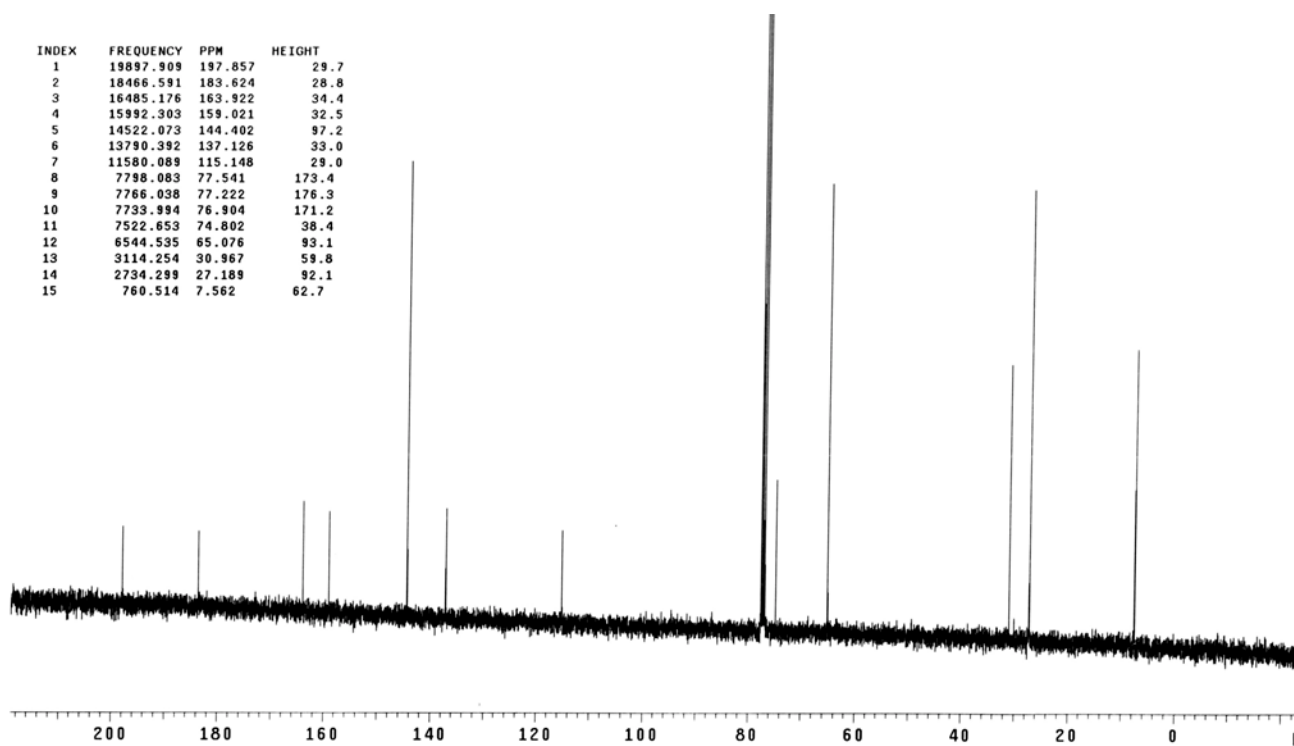
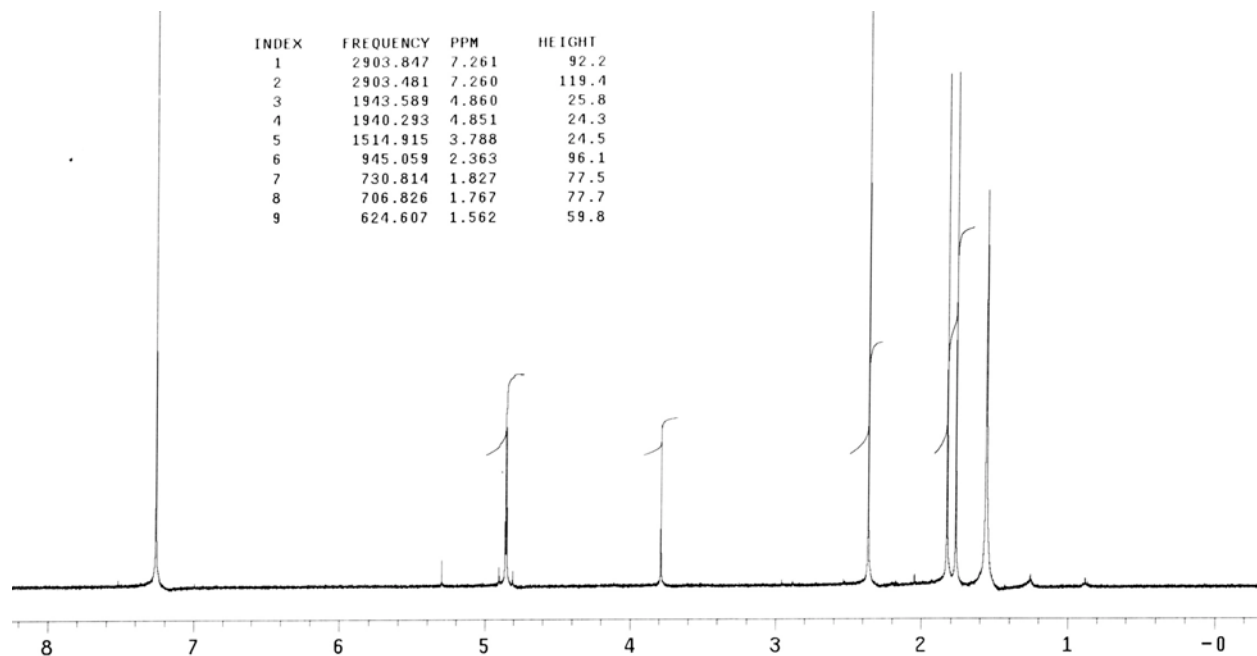


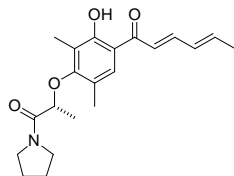
Amide 14: pale yellow oil $R_f=0.50$ EtOAc (100%); ^1H NMR (CDCl_3 , 400 MHz) δ 7.40 (m, 1H), 6.90 (br s, OH), 6.31 (d, 1H, $J = 15.0$ Hz), 6.31–6.20 (m, 2H), 4.91 (d, 1H, $J = 15.0$ Hz), 4.52 (d, 1H, $J = 15.0$ Hz), 3.62 (s, 1H), 3.54 (t, 2H, $J = 7.0$ Hz), 3.35 (m, 1H), 3.27 (m, 1H), 2.05 (m, 2H), 1.89–1.85 (m, 2H), 1.88 (d, 3H, $J = 6.2$ Hz), 1.84 (s, 3H), 1.56 (s, 3H); ^{13}C NMR (CDCl_3 , 100.6 MHz) δ 191.9, 191.5, 171.8, 167.3, 146.8, 143.3, 130.7, 123.0, 115.7, 70.5, 70.4, 62.7, 61.4, 46.7, 45.3, 26.7, 26.3, 24.0, 19.2, 10.2; IR (CHCl_3 , $\lambda\text{-max cm}^{-1}$) 3280, 3020, 1645, 1217; HRMS(EI) calcd for $\text{C}_{20}\text{H}_{25}\text{NO}_6$ 375.1682, found: 375.1692.



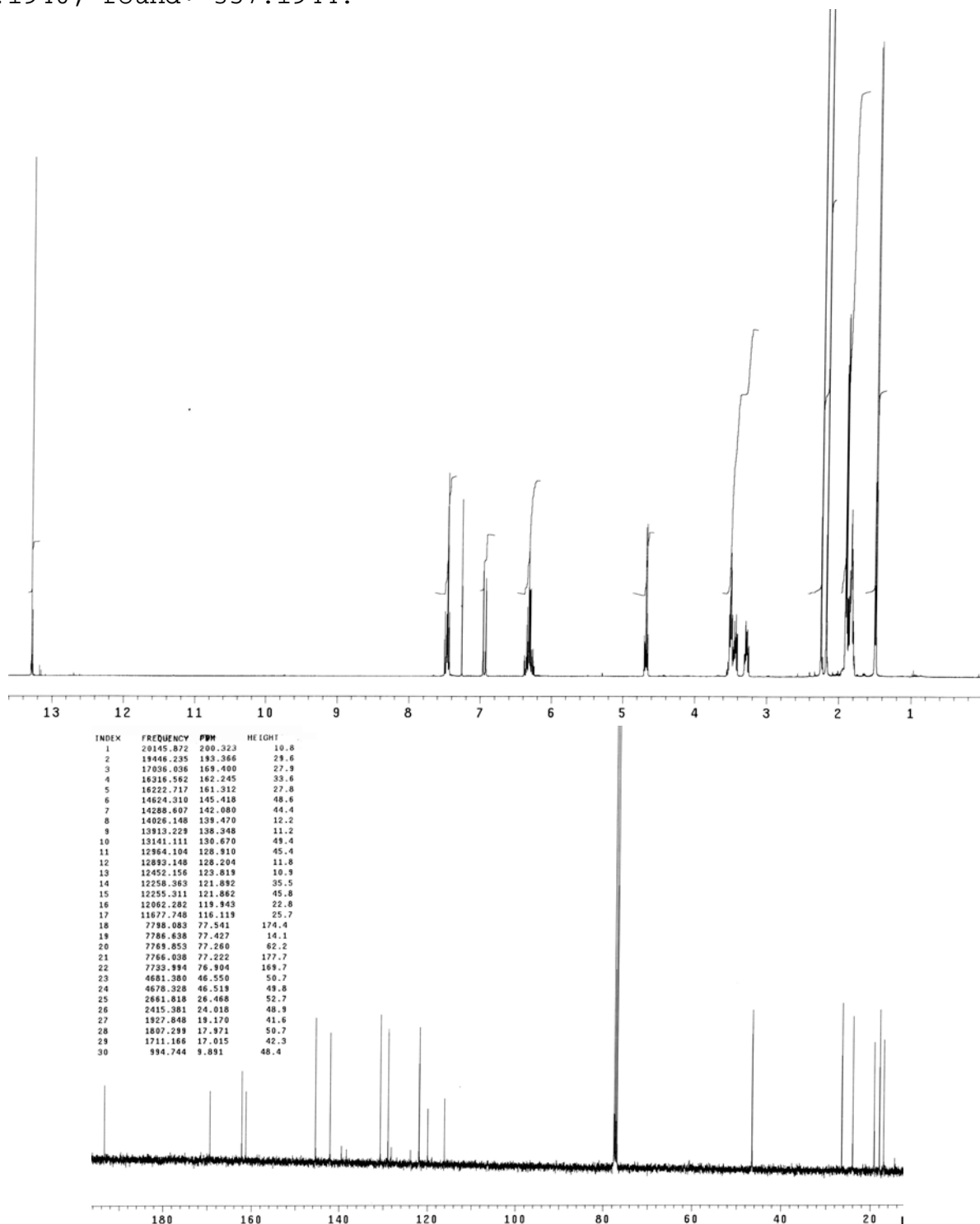


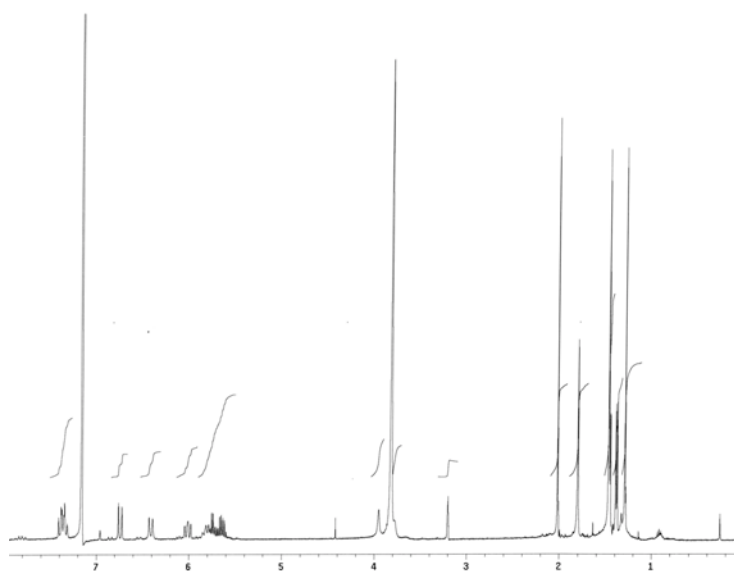
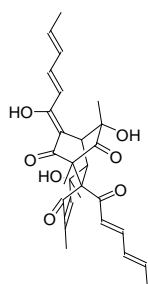
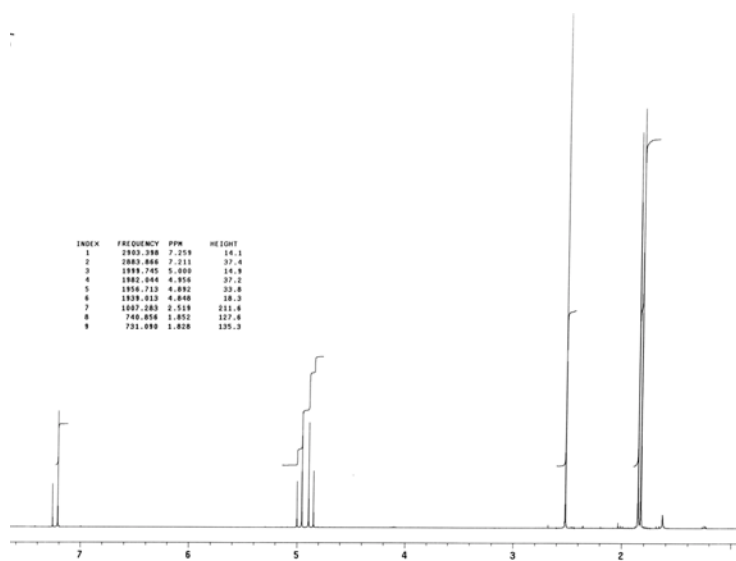
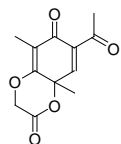
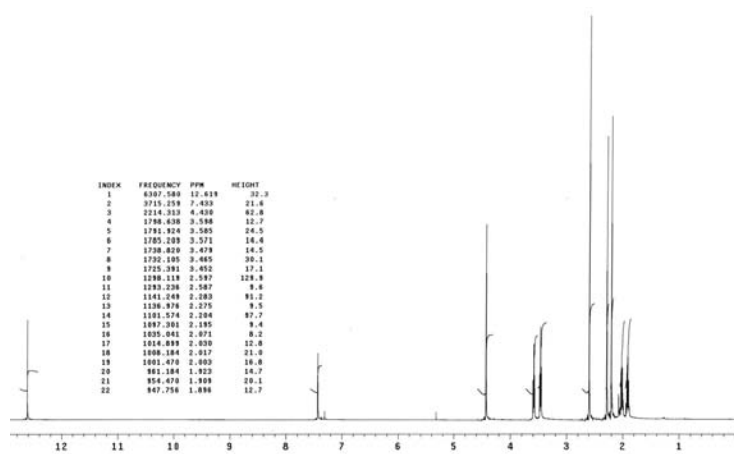
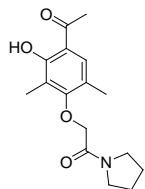
epoxide 16 colorless solid: m.p. 165-167 °C; R_f=0.47 Hexanes/EtOAc (1/1); ¹H NMR (CDCl₃, 400 MHz) δ 4.85 (s, 2H), 3.79 (s, 1H), 2.34 (s, 3H), 1.81 (d, 3H, J = 5.2 Hz), 1.75 (s, 3H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 200.0, 189.2, 163.9, 157.0, 110.8, 76.6, 64.5, 61.2, 58.6, 28.2, 25.7, 8.1; IR (CHCl₃, λ_{max} cm⁻¹) 3022, 1776, 1730, 1642, 1294; HRMS(EI) calcd for C₁₂H₁₂O₆ 252.0634, found: 252.0636.

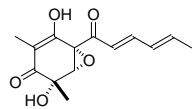




Lactic Acid Derivative 17 yellow oil Rf=0.55
 $\text{CH}_2\text{Cl}_2/\text{acetone}$ (12/1); $[\alpha]_D = +35.1^\circ$ (c 1.0, CHCl_3); ^1H
 NMR (CDCl_3 , 400 MHz) δ 13.30 (s, OH), 7.46 (m, 2H), 6.95
 (d, 1H, $J = 15.0$ Hz), 6.32 (m, 2H), 4.66 (q, 1H, $J =$
 6.8 Hz), 3.50 (m, 2H), 3.42 (m, 1H), 3.29 (m, 1H), 2.24
 (s, 3H), 2.17 (s, 3H), 1.90 (d, 3H, $J = 5.8$ Hz), 1.88–1.59 (m, 4H),
 1.49 (d, 3H, $J = 6.8$ Hz); ^{13}C NMR (CDCl_3 , 100.6 MHz) δ 193.4, 169.4,
 162.2, 161.3, 145.4, 142.1, 130.7, 128.9, 121.9, 121.8, 119.9, 116.1,
 77.3, 46.6, 46.5, 26.5, 24.0, 19.2, 18.0, 17.0, 9.9; IR (CHCl_3 , $\lambda\text{-max}$
 cm^{-1}) 3685, 3027, 2983, 1644, 1622, 1360; HRMS(EI) calcd for $\text{C}_{21}\text{H}_{27}\text{NO}_4$
 357.1940, found: 357.1944.







synthetic epoxysorbicillinol (**1**) yellow amorphous powder
 $R_f=0.16$ $\text{CH}_2\text{Cl}_2/\text{methanol}$ (7/1); ^1H NMR (CDCl_3 , 400 MHz) δ 10.16 (s, enolic OH), 7.58 (m, 1H), 6.45 (m, 1H), 6.30 (m, 2H), 3.98 (s, H6 and OH), 1.95 (d, 3H, $J = 6.8$ Hz), 1.81 (s, 3H), 1.35 (s, 3H); ^1H NMR (CD_3OD , 400 MHz) δ 7.29 (m, 1H), 6.66–6.27 (m, 3H), 3.56 (s, 1H), 1.86 (d, 3H, $J = 5.9$ Hz), 1.64 (s, 3H), 1.46 (s, 3H); ^{13}C NMR (CD_3OD , 100.6 MHz) δ 194.7, 189 (evasive)^{1d}, 173 (evasive)^{1d}, 147.4, 144.5, 131.7, 124.7, 107.8, 70.6, 64.0, 62.9, 26.3, 19.2, 8.1; IR (CHCl_3 , ν_{max} cm^{-1}) 3420, 3009, 1675, 1390, 800; HRMS(CI) calcd for $\text{C}_{14}\text{H}_{16}\text{O}_5$ ($\text{M}+\text{H}$)⁺ 265.0998, found: 265.1007.

